

**ORIGINAL RESEARCH PAPER**

**EVALUATION OF NUTRITIONAL QUALITY OF THE COMMON CARP  
(*CYPRINUS CARPIO*) ENRICHED IN FATTY ACIDS**

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The knowledge of the feed requirements of fish species allows developing well-balanced feed formulations for successful intensive culture. The target of this study was to investigate the effect of supplementing the fish fodder with different types of oils, on fish growing and meat quality. The experiments were conducted on common carp using olive, soy and fish oil preparations. The results of these experiments showed that increasing the content of fats in fish diet allowed a faster growth of common carp and better weight increasing results. The fatty acids and aminoacids profiles of the fish meat highly depended on the type of oil used in the feeding experiment. According to our results, out of the tested variants the fish oil based fodder is most favorable for fish feeding, allowing the accumulation of large quantities of highly unsaturated fatty acids in meat.

**Keywords:** common carp, fodder, fish oil, olive oil, soy oil, fatty acids, aminoacids

### **Introduction**

Common carp (*Cyprinus carpio* L.) is the main aquaculture species in many European, Asian and Latin American countries.

In aquatic habitat, the fish take the energy for growing and reproduction from the available plankton, periphyton, nuston, benthose, nekton and plants (Narejo *et al.*, 2010). The successful fish culture requires the use of efficient feed with optimum composition. In the last years many studied have been focused on the fish diet allowing development of new balanced commercial products which promote optimal growth of healthy fish. In aquaculture fish nutrition is critical because the fodder contributes with 40-50% to the total production costs. Development of new

feed formulations for certain fish species supports the industrial aquaculture, which must expand to meet the growing demand for safe and high quality fish products at affordable prices.

When fish are raised in closed systems or in high density ponds, they must be provided with a high quality diet, complete and nutritionally balanced, allowing the rapid fish growth and good health. The complete fish feed provides all essential nutritional compounds, such as proteins, carbohydrates, fats, vitamins and minerals (Gangadhara *et al.*, 1997; Cho *et al.*, 2005). Most of the fish feeds available on the market have the following composition: proteins (18-50%), lipids (10-25%), carbohydrates (15-20%), ash (<8.5%), phosphor (<1.5%), moisture (<10%) and traces of vitamins and minerals.

On the other hand, the incomplete fish feed/diets do not contain the whole range of vitamins or mineral and are designed to act as supplements to the natural feeding available in the open fish farming systems.

The effect of fish and vegetable oils on fish nutrition and health is not enough studied. Some authors reported that oil supplemented diets improve the lipid contents in case of *Pseudoplatystoma coruscans* (Martino *et al.*, 2002), *Labeo rohita* (Satpathy *et al.*, 2003), *Scophthalmus maximus* L. (Cho *et al.*, 2005), *Epinephelus coioides* (Luo *et al.*, 2005), *Sparus latus* (Hu *et al.*, 2007), *Epinephelus malabaricus* (Tuan and Williams, 2007) and *Sander lucioperca* (Schulz *et al.*, 2008).

The contents of different types of fatty acids in fish and vegetable oils are very different: unlike vegetable oil, fish oil is rich in highly unsaturated fatty acids. Due to these differences, it is expected to involve different metabolic pathways and to have a different impact on fish nutrition and health. The aim of the present study was to investigate the effect of feeding the common carp with fodder supplemented with olive, soy and fish oil. After the feeding experiment, the fish meat was checked in terms of fatty acid and aminoacid profiles.

## Materials and methods

### Materials

The experiment was conducted in the autumn of 2011 (from 14th of September to 7th of November) on common carp (*Cyprinus carpio*).

The cyprinids obtained through natural reproduction system in the spring of 2011, were transferred from the Base of Research – Development Brates, Galati into water basins where the fish was fed for 54 days with different nutritional supplements.

### Fish feeding experiments

The feeding experiments were carried out in basins of the National Research and Development Institute for Aquatic Ecology, Fishing and Aquaculture, Galati. The feed ratio was 5% from the total mass of biologic mate. The water parameters (O<sub>2</sub>, pH, temperature, NH<sub>4</sub><sup>+</sup>, NO<sub>2</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>) were monitored twice a day and were checked to be in agreement with the 161/2006 recommendation for the second order quality.

Four different types of fodder were used for feeding the summer-aged common carp:

- experiment I (codified with M) - the feeding was performed using the commercial by available fodder CLASSIC K, which is specially developed for common carp feeding in agreement with *AminoBalance* and *LipoBalance* concepts to ensure the optimum equilibrium of amino acids and lipids profiles;
- experiment II (codified with FC1) - the feeding was performed with CLASSIC K fodder supplemented with 5% olive oil, having 12.8% saturated fatty acids, 71% monounsaturated fatty acids and 7.8% polyunsaturated fatty acids;
- experiment III (codified with FC2) - the feeding was performed with CLASSIC K fodder supplemented with 5% soy oil provided by SC 22E-Prod SRL, Alexandria, Romania;
- experiment IV (codified with FC3) - the feeding was performed with CLASSIC K fodder supplemented with 5% fish oil obtained from cod liver, with the following fatty acids composition: 1.6 g saturated fatty acids/10 ml, 4.6 g monounsaturated fatty acids/10 ml, 3 g polyunsaturated fatty acids/10 ml, 0.7 g eicosapentaenoic acid (EPA)/10 ml and 0.9 g docosahexaenoic acid (DHA)/10 ml.

According to the producers, the CLASSIC K fodder has the following ingredients: whole soy flour, corn, fish flour, wheat, sunflower grist, fish oil, soy oil, rapeseed oil and butylhydroxytoluene. The fodder supplementation with oils was carried out in the laboratories of National Research and Development Institute for Aquatic Ecology, Fishing and Aquaculture, Galati, using a laboratory extruder.

Ten specimens from the four different experiments were randomly selected for biometric and physico-chemical analysis.

#### ***Physical and chemical analysis***

The composition of the fodder and of the fish before and after feeding in the basins was checked.

The proximate composition was determined as follows: the moisture, protein and ash contents were determined according to standard AOAC (1995) methods; the fat content by extraction with ether through Soxhlet method. The total glucides content was calculated by subtracting the protein, fat, ash and moisture content from the total weight of the samples.

pH measurements were made according to AOAC (1984). Ten grams of sample were homogenized with 90 ml distilled water for 2 min using the Braun mixer. The obtained mixture was filtered and the pH of the filtrate determined by means of a Hanna digital pH-meter.

The oxidation degree of the fish meat was estimated by determining the thiobarbituric acid (TBA) index (mg malondialdehyde/kg sample) according to Ionescu *et al* (1992).

### ***Fatty acids profiling***

The fatty acids profiles of both fodder and fish meat was determined by means of gas chromatography (GC).

In order to extract the lipids, the homogenized samples were dried for 1 h at 105°C. Prior to analysis, the fat was subjected to saponification and the free fatty acids were converted to methyl esters.

GC analyses were performed on Perkin Elmer CLARUS 500 equipped with flame ionization detector (FID) and with DB-23 capillary column (length 60 m, diameter 0.25 mm, film thickness 0.25 µm).

The injector temperature was 250°C, and samples were injected manually (1 µl). The temperature increased from 180°C to 280°C at a rate of 5°C/minut. The hydrogen was used as carrier gas at a flow rate of 35 cm/s, 180°C. The detector temperature was 260°C.

The standard used was Supelco-Supelco ® 37 Component FAME Mix.

### ***Amino acids profiling***

The amino acids profile of the fish meat was determined by means of High-Performance Liquid Chromatography (HPLC). The samples were prepared for analyses by the oxidation of sulphur-containing amino acids and acidic hydrolysis of the peptide bonds.

The HPLC Surveyor Plus (Thermo Electro) was used for analyses and the amino acids were separated on a Hypersil BDS C18 column (250 x 4.6 mm, 5 µm) in conjunction with a gradient elution. Eluent A was sodium phosphate buffer pH 7 and eluent B was a mixture of acetonitrile:methanol:water (45: 45: 10). The volume of sample injected was 25 µl, the flow rate was 1.0 ml/min and the column temperature was set at 25°C. The emission wavelength was 336 nm.

### ***Statistical analysis***

Statistical analysis was carried out by means of Excel tools. All analyses were carried out in triplicate. The average values are reported together with standard deviations.

## **Results and discussion**

### ***Chemical analyses of fodder***

The composition of the fodders used for fish feeding is presented in Table 1. The protein content of the fodder samples supplemented with different types of oil decreased; the lowest value (39.59%) was obtained in case of the fodder supplemented with 5% soy oil.

In case of the fodders with olive and fish oils, the increase of the fat content was less than 5% with respect to FM. We can explain this difference by the strong binding of the fat to other fodder components, so as to be impossible to extract it by solvents after extrusion.

The energy value of the FM sample was comparable with that indicated by the producer, but lower with respect to the samples supplement with different types of oils (Table 1).

**Table 1.** Chemical composition of the fodders

	Fodder sample			
	FM	FC1	FC2	FC3
Moisture, g%	5.55±0.102	5.11±0.049	4.94±0.007	5.16±0.015
Proteins, g%	45.52±0.045	41.47±0.044	39.59±0.040	40.19±0.067
Fats, g%	10.67±0.010	13.38±0.058	15.51±0.023	13.34±0.015
Ash, g%	5.68±0.070	5.73±0.055	5.55±0.027	4.88±0.018
Total carbohydrates*, g%	32.58	34.31	34.41	36.43
TBA index, mg MDA**/kg	2.81±0.004	2.73±0.001	4.40±0.006	3.81±0.004
Energy value, kcal/100 g ***	343.89	353.64	364.88	351.17

\* Total carbohydrates and other chemical components estimated by subtracting the moisture, proteins, fats and ash from the total weight;

\*\*MDA-malondialdehyde;

\*\*\*calories conversion factors used: for proteins 4.3 kcal/g, for lipids 9.0 kcal/g; for carbohydrates 1.6 kcal/g.

The composition of the fodders used in our experiment is quite different with respect to those indicated by Kukačka *et al.* (2009) that have protein contents of 29.5-33.8% and fat contents ranging from 9.1 to 19%. Moreover, Manjappa *et al.* (2005) used for carp feeding fodder with the following proximate composition: proteins 23.85-24.84%, fats 6.85-13.01% and ash 12.98-13.38%.

The addition of soy and fish oils caused the significant increase of the compounds reacting with TBA. The high values of the TBA index in case of these samples (4.4 and 3.81 mg MDA/kg in case of FC2 and FC3, respectively) give indications about the reduced oxidative stability which requires special storing conditions such as low temperature, absence of light, low relative humidity and the use of well-sealed packages.

The profiles of the fatty acids of the fodders used in our experiments were comparable (Table 2). The level of saturated fatty acids varied in the following order: FM>FC3>FC1>FC2. The addition of soy oil caused the most important decrease of the saturated fatty acids, mostly as a consequence of the reduced content of palmitic acid. The addition of vegetable and fish oils caused the increase of the monounsaturated fatty acids (MUFA) levels, mainly of oleic acid cis omega 9. The presence of miristoleic acid was registered only in the case of the fodder with soy oil.

The level of polyunsaturated fatty acids (PUFA) with 2 or 3 double bonds decreased in the following order: FC2>FM4>FC1>FC3 (Table 2). The soy oil had important quantities of cis linoleic acid, while olive and fish oils had lower amounts of cis linoleic acid (C18:2n-6) and  $\alpha$  linolenic acid (C18:3n-3) (Table 2).

The fish oil had high levels of highly unsaturated fatty acids (HUFA): 4.95 g EPA /100 g fats and 5.09 g DHA /100 g fats. The fodder FC3 had the highest n-3/n-6 value (0.58), while the FC2 had the lowest values (0.44).

**Table 2.** The profiles of the fatty acids in the fodder samples

Fatty acid	Fodder sample			
	FC1, g/100 g fats	FC2, g/100 g fats	FC3, g/100 g fats	FM, g/100 g fats
Miristic acid (14:0)	2.55	2.40	2.80	3.13
Palmitic acid (C16:0)	13.44	11.70	13.68	14.55
Heptadecanoic acid (C17:0)	0.44	0.40	0.51	0.51
Stearic acid (C18:0)	2.58	2.53	2.50	2.49
<b>Total saturated fatty acids</b>	<b>19.01</b>	<b>17.03</b>	<b>19.49</b>	<b>20.68</b>
Miristoleic acid (C14:1)	0.00	0.12	0.00	0.12
Palmitoleic acid (C16:1)	3.11	2.90	3.67	3.48
Heptadecenoic acid (C17:1)	0.33	0.32	0.38	0.40
Cis-Oleic acid (C18:1:n-9)	29.30	25.99	25.63	24.21
Oleic acid (C18:1:n-7)	2.18	2.14	2.45	2.19
Eicosanoic acid (C20:1:n-9)	1.61	2.05	2.36	1.94
Erucic acid (C22: (1:n-9)	1.51	1.85	2.06	1.84
Nervonic acid (C24:1n-9)	0.59	0.65	0.72	0.77
<b>Total MUFA</b>	<b>38.63</b>	<b>36.02</b>	<b>37.27</b>	<b>34.95</b>
Cis- Linoleic acid (C18:2:n-6)	26.68	29.67	24.81	26.99
Conjugate linoleic acid	1.25	1.24	1.43	1.34
Eicosadienoic acid (C20:2:n-6)	0.25	0.28	0.32	0.34
$\gamma$ Linolenic acid (C18:3:n-6)	0.11	0.10	0.12	0.00
$\alpha$ Linolenic acid (C18:3:n-3)	4.49	5.40	4.16	4.77
Eicosatrienoic acid (C20:3:n-3)	0.30	0.22	0.31	0.29
<b>Total PUFA (2 or 3 double bonds)</b>	<b>33.08</b>	<b>36.91</b>	<b>31.15</b>	<b>33.73</b>
Octadecatetraenoic acid (C18:4n-3)	0.15	0.20	0.25	0.14
Arachidonic acid (C20:4:n-6)	0.10	0.14	0.00	0.13
Eicosapentaenoic acid (C20:5:n-3)	3.83	3.76	4.95	4.47
Docosahexaenoic acid (C22:6:n-3)	3.91	3.70	5.09	4.79
<b>Total HUFA (&gt;4 double bonds)</b>	<b>7.99</b>	<b>7.8</b>	<b>10.29</b>	<b>9.53</b>
Other fatty acids	1.29	2.26	1.82	1.09
Total $\omega$ -6 fatty acids	27.14	30.19	25.25	27.46
Total $\omega$ -3 fatty acids	12.68	13.23	14.76	14.46
n-3/n-6	0.46	0.44	0.58	0.53

Kukačka *et al* (2009) reported the use of fodder supplemented with 6% fish oil (sample codified Ro6) for common carp feeding. The fatty acids profiles of Ro6 and FC3 are very different. In case of Ro6, only 18 different fatty acids were identified while in our case 22 fatty acids were found. Compared to FC3, higher levels of saturated fatty acids (27.95%) and PUFA (48.41%) and lower levels of MUFA (23.64%) were reported by Kukačka *et al* (2009) for Ro6. Moreover, the n-

3/n-6 ratio was 1.15 in the case of Ro6 and only 0.58 in the case of FC3. These differences in terms of fatty acids are mainly due to the particular composition of the fodders. The Ro6 had the following composition: fish flour (16%), rapeseed flour (12%), soy flour (12%), Vitex (10%), whey (3%); wheat (4%), wheat flour (30%), lecithin (3%), and Aminovitan (2%) (Kukačka *et al.*, 2009).

#### ***Analyses of the fish before differential feeding***

The common carp used for differential feeding experiments had the following biometric parameters: length of 129.37±18.45 mm, diameter of 110.88±8.04 mm and weight of 59±17.07 g. The statistical analyses indicated good linear correlation coefficients between weight and length ( $r = 0.976$ ) and between length and diameter ( $r = 0.931$ ).

The ponderal anatomy parameters of the common carp are presented in Table 3. The amount of fish meat before differential feeding experiments correlates well with the fish weight ( $r = 0.840$ ) and only lower linear correlation coefficients were identified in case of the viscera and bones ( $r = 0.494$ ).

**Table 3.** Ponderal anatomy of fish before and after 54 days of differential feeding experiments

Fish sample	Meat, %	Head, %	Viscera, %	Scales, %	Skin, %	Bones, %	Flippers, %	Blood, %
Initial	42.70	27.82	11.65	3.42	5.72	5.98	2.33	0.31
FC1	41.55	21.12	13.20	3.16	8.68	8.48	3.27	0.54
FC2	42.05	22.16	11.09	3.90	9.21	7.17	3.92	0.50
FC3	42.33	22.17	12.68	3.21	7.02	8.35	3.85	0.39
FM4	41.44	22.27	13.98	4.07	6.55	7.27	3.92	0.51

The proximate composition of fish meat is presented in Table 4. Before differential feeding experiments fish meat had high moisture contents, medium protein contents and low fat contents, and therefore had slow energy value of 70.41 kcal/100g.

**Table 4.** Proximate composition of the fish meat before and after 54 days of differential feeding experiments

	Fish sample				
	Initial	FC1	FC2	FC3	FM
Moisture, %	82.98±0.20	81.08±0.26	79.98±0.13	79.94±0.04	79.58±0.15
Proteins, %	15.16±0.24	15.34±0.17	16.6±0.20	17.14±0.07	14.88±0.07
Fats, %	0.33±0.013	0.53±0.009	0.34±0.003	0.32±0.006	0.56±0.001
Ash, %	1.05±0.034	0.91±0.011	1.05±0.002	1.09±0.001	1.01±0.05
Glucides, %		2.14	2.03	1.55	3.97
pH	6.70	6.63	6.70	6.59	6.28
TBA index, mg					
MDA/kg	0.65±0.04	0.51±0.006	1.45±0.012	0.42±0.004	0.69±0.008
Energy value, kcal/100g	70.41	75.19	78.84	80.98	71.77

### *Analyses of fish after differential feeding*

Fish fed with fodder supplemented with fish oil had the highest biometric parameters (Table 5). The fodder supplemented with olive oil was less efficient, the effect on common carp growing being comparable with that of FM. The efficiency of fish growing in case of the feeding with FC 2 and FC3 fodders might be due to the efficient use of available proteins from diet and to the appropriate lipids satisfying the energetic requirements (Caballero, *et al.*, 1999; Satpathy, *et al.*, 2003). The carp proteins requirements vary with the age from 25% to 35% (Hossain, *et al.*, 1997). Steffens (1996) suggested that the increase of diet energetic density can be a strategy for proteins saving and limiting the ammonium production in case of different fish species, including common carp.

According to Manjappa *et al.* (2002) the addition of 3% sardine oil in common carp fodder as well as a diet with 10% fish flour allows an appropriate growth of the fish. Gangadhara *et al.* (1997) showed, in the case of the Indian carp rohu (*Labeo rohita*), that diets with 25% proteins and 9% fats or with 30% proteins and 6% fats allowed comparable results in terms of growing.

**Table 5.** Biometric parameters of the common carp after differential feeding

Fish sample	Length, mm	Diameter, mm	Weight, g	Weight gain, g*	Length gain, mm*
FC1	212.0	185.0	297.0	238.0	82.625
FC2	232.0	209.0	343.0	284.0	102.625
FC3	267.0	214.0	491.0	432.0	146.625
FM	182.5	167.5	162.5	103.5	53.125

\*weight and length gain were established in direct relation with the mean values of the samples before differential feeding

Concerning the ponderal anatomy of the fish differentially fed, the results presented in Table 3 indicate that there are no significant differences within samples. On the other hand, the proximate composition varied with the type of fodder (Table 4).

The fish fed with fodder supplemented with fish and soy oil had 1.1-1.98% lower moisture contents with respect to the FC1 and FM. The dry matter increase is mainly related to the advanced protein synthesis (Fafioye *et al.*, 2005); the protein levels in the FC2 and FC3 fish samples were 16.6% and 17.14, respectively. Our results concerning the increase of protein contents in the case of supplementing the fish diets with oil are in agreement with the observations of Gangadhara *et al.* (1997) for *Labeo rohita*, of Bazaz and Keshavanath (1993) for *Tor khudree* and of Cho *et al.* (2005) for *Scophthalmus maximus* L.

The fat content of the FC2 and FC3 fish samples is comparable with that of the fish before starting the differential feeding experiment. The highest fat contents of 0.56±0.001% and 0.53±0.009% were identified in case of the FM and FC1 fish samples, respectively (Table 4). Our results are lower compared to those reported by Steffens and Wirth, (2007) for the carp naturally fed (6.8%) and by Fajomová *et al.* (2003) for the carp fed with diets supplemented with 7% cereals. Steffens and



Wirth (2007) reported 3.4 g fats/100g d.w. when supplementing the fish diets with cereals. These variations in the fat content can be explained by the differences in the fish weight, diet, growing conditions and samples preparation procedure.

On the other hand, the increase of oil in the diet causes the increase in lipase activity in the hepatopancreas of *Tor khudree*, *Labeo rohita* and *Dicentrarchus labrax* (Bazaz and Keshavanath, 1993; Gangadhara et al., 1997; Peres and Oliva-Teles, 1999).

Analyzing the results in Table 4, one can see that the fodder FC2 caused the accumulation of high levels of compounds reacting with 2-thiobarbituric acid that can significantly influence fish health and consumer health.

Fodder supplementation with oils caused relative by small increases in the energetic value compared to the control sample (Table 4). The highest energetic value was obtained in the case of the FC3 fish sample (80.98 kcal/100g) which is ~13% higher compared to the control sample.

### **Fatty acids profiling**

The fatty acids profiles of the fish meat samples are presented in Table 6. Feeding the fish with fodder supplemented with 5% fish, soy or olive oil caused the reduction of the total amount of saturated fatty acids compared to the control sample. The lowest level of palmitic acid (14.35 g/100 g fats) was registered in the case of FC2 fish sample (Table 6).

Concerning the content of MUFA, the FC1 fodder caused the accumulation in the fish meat of 43.97 g MUFA/100 g fats, which is comparable to the control sample (41.85 g/100 g fats) and much higher compared to the FC2 (38.87 g/100 g fats). The contents of palmitoleic (C16:1) and cis-oleic acids (C18:1n-9) are lower in the case of FC2 and FC3 fish samples compared to the FC1 and FM (Table 6). The presence of fish oil in the fodder was the origin of doubling the content of docosanoic acid (C22:1:n-11) with respect to the control sample. A similar trend was observed also in the case of eicosanoic acid (C20:1:n-9), but at higher levels. The highest level of nervonic acid (C24:1n-9) was found in the FC3 fish sample.

The differential fish feeding also reflected in the PUFA profile. The addition of soy oil in fish diet was reflected in the significant increase of PUFA to 32.73 g/100 g fats, while the fish oil induced higher synthesis of conjugated linoleic acid (0.93 g/100 g fats) and eicosatrienoic acid (C20:3:n-3) (3.49 g/100 g fats).

The diet with fish oil favored the highest accumulation of HUFA (9.59 g/100 g fats) in fish, while the other samples had HUFA content ranging from 4.79 to 5.77 g/100 g fats. The highest amounts of HUFA in FC3 fish sample are 5.28 g DHA/100 g fats 2.52 g EPA/100 g fats, and 0.8 g DPA/100 g fats. On the other hand, high amounts of octadecatetraenic acid (C18:4n-3) and DHA were found in the fish samples fed with soy oil (Table 6). The high content of polyunsaturated fatty acids has beneficial health effects (Steffens, 1996).

According to Ikeda et al., (2011), the source of vegetable oil used for fish diet supplements does not affect the growth and thermal tolerance of the fish. The common carp with high lipid deposits has an improved cold resistance.

**Table 6.** The profiles of fatty acids in the fodder samples

Fatty acid	Fish sample			
	FC1, g/100g fats	FC2, g/100g fats	FC3, g/100g fats	FM, g/100g fats
Myristic acid (C14:0)	1.47	0.94	2.09	1.28
Pentadecanoic acid (C15:0)	0.24	0.19	0.31	0.22
Palmitic acid (C16:0)	16.48	14.35	15.09	17.73
Heptadecanoic acid (C17:0)	0.28	0.27	0.4	0.22
Stearic acid (C18:0)	4.38	5.05	4.69	4.97
<b>Total saturated fatty acids</b>	<b>22.85</b>	<b>20.8</b>	<b>22.58</b>	<b>24.42</b>
Myristoleic acid (C14:1)	0.13	0.1	0.2	0.1
Pentadecenoic acid (C15:1)	0.52	0.53	0.6	0.49
Palmitoleic acid (C16:1)	5.56	3.95	5.07	6.01
Heptadecenoic acid (C17:1)	0.29	0.24	0.34	0.26
Trans-oleic acid (C18:1:n-11)	0.07	0.11	0.11	0.11
Cis-oleic acid (C18:1:n-9)	32.25	28.59	25.6	30.14
Oleic acid (C18:1:n-7)	2.33	2.33	2.69	2.36
Docosanoic acid (C22: 1:n-11)	0.43	0.6	1.31	0.24
Erucic acid (C22:1:n-9)	0.06	0.06	0.15	0.08
Eicosanoic acid (C20: 1:n-9)	1.81	1.79	2.63	1.54
Nervonic acid (C24:1:n-9)	0.52	0.57	0.86	0.52
<b>Total monounsaturated fatty acids</b>	<b>43.97</b>	<b>38.87</b>	<b>39.56</b>	<b>41.85</b>
Cis-linoleic acid (C18:2:n-6)	18.87	24.78	17.03	19
Conjugated linoleic acid (CLA)	0.62	0.56	0.93	0.42
Eicosadienoic acid (C20:2:n-6)	0.88	0.72	0.81	0.95
$\gamma$ -linoleic acid (C18:3:n-6)	0.39	0.32	0.42	0.48
$\alpha$ -linolenic acid (C18:3:n-3)	2.36	3.18	2.05	1.84
Eicosatrienoic acid (C20:3:n-6)	0.74	0.73	0.93	0.87
Eicosatrienoic acid (C20:3:n-3)	2.23	2.44	3.49	3.13
<b>Total polyunsaturated fatty acids (PUFA, &gt; 2 double bonds)</b>	<b>26.09</b>	<b>32.73</b>	<b>25.66</b>	<b>26.69</b>
Arachidonic acid (C20:4:n-6)	0.12	0.1	0.13	0.09
Eicosapentaenoic acid (C20:5:n-3) (EPA)	1	1.16	2.52	0.82
Octadecatetraenoic acid (C18:4:n-3)	0.31	0.63	0.49	0.26
Docosatetraenoic acid (C22:4:n-6)	0.22	0.26	0.37	0.31
Docosapentaenoic acid (C22:5:n-3) (DPA)	0.52	0.47	0.8	0.75
Docosahexaenoic acid (C22:6:n-3) (DHA)	2.7	3.15	5.28	2.56
<b>Total highly unsaturated fatty acids (HUFA, &gt; 4 double bonds)</b>	<b>4.87</b>	<b>5.77</b>	<b>9.59</b>	<b>4.79</b>
Other fatty acids	2.22	1.83	2.63	2.26
Total $\omega$ -6 fatty acids	21.22	26.91	19.69	21.7
Total $\omega$ -3 fatty acids	9.12	11.03	14.63	9.36

**Amino acids profiling of the fish meat**

In Table 7 is presented the amino acids profile of the analyzed fish meat samples. Out of the aminoacids known to be present in fish meat, histidine and hydroxyproline were not identified and quantified.

The acid treatment used to hydrolyze the proteins to aminoacids is known to affect totally the tryptophan residues and partially the sulfur containing aminoacids.

There are no significant differences between fish samples in terms of aminoacids contents (Table 7). The fish sample FC1 had lower lysine and higher tyrosine and isoleucine contents compared to the other samples. The control sample had the highest contents of lysine, phenylalanine, isoleucine and leucine and the lowest proline contents.

**Table 7.** The amino acids profile of fish samples fed with different types of fodder

Aminoacid	Fish sample				Common carp, g/100 g proteins
	FC1, g/100 g proteins	FC2, g/100 g proteins	FC3, g/100 g proteins	FM4, g/100 g proteins	
Aspartic acid	6.98	7.22	7.11	7.96	9.60
Glutamic acid	13.36	13.33	12.93	13.94	14.40
Serine	4.87	4.52	4.33	4.11	4.40
Glicine	6.08	5.63	5.60	4.20	7.00
Treonine	4.02	3.83	3.72	3.98	4.70
Arginine	7.99	7.56	7.27	7.58	6.90
Alanine	6.59	6.37	6.19	6.26	7.20
Tyrosine	8.24	6.6	6.21	7.23	3.30
Valine	2.26	2.23	2.16	1.77	5.90
Phenylalanine	3.72	3.57	3.43	4.24	4.00
Isoleucine	3.9	3.62	3.60	4.30	4.60
Leucine	9.18	8.49	8.32	8.66	8.30
Lysine	12.48	14.28	14.45	15.25	8.20
Proline	2.13	2.41	2.86	1.40	4.80
Cystine	0.44	0.39	0.27	0.49	0.90
Methionine	1.69	1.43	1.45	1.40	3.40
Other aminoacids*	6.07	8.52	10.1	7.23	2.40

\* estimated by subtracting the total amount of aminoacids quantified from 100 g proteins

The fish diet and other growing conditions influence the protein composition of the common carp. Comparing the profile of the aminoacids of the fish samples differentially fed to that of the wild common carp (Table 7), one can see that the levels of aspartic acid, glutamic acid, glycine, alanine, valine, proline, cystine and methionine were higher, while the levels of tyrosine, lysine and other aminoacids are lower in the case of the wild fish. The ratio essential/nonessential aminoacids

was significantly higher in the case of the wild common fish compared to the experimental ones.

### Conclusions

The fat rich diets allowed a faster growth of the common carp due to a more efficient use of the fodder. The best fish weight growing results were obtained in case of feeding the fish with fodder supplemented with fish oil. The use of the fodder supplemented with fish and soy oil allowed obtaining fish samples with higher dry matter, protein and ash contents compared to the case when the olive oil was used. The fat content of FC1 fish sample was comparable to the control and higher than the FC2 and FC3 fish samples.

The supplementation of the fish diets with lipids reflected on the fatty acids profile of the fish meat. The fodder with fish oil favored the accumulation of HUFA, especially EPA, DPA and DHA. The fodder supplemented with oils of vegetable origin did not influence the ratio n-3/n-6 fatty acids in the fish meat with respect to the control sample.

The different fodder used in the experiment also caused variations in the aminoacids profile of the fish meat; the ratio essential/nonessential aminoacids is lower compared to the wild common carp.

Taking into account the results obtained we may conclude that the best fodder is CLASSIC K supplemented with 5% fish oil.

### References

- AOAC. 1984. Official methods of analysis (11th ed.). Washington, DC: Association of Official Analytical Chemists.
- AOAC. 1995. Official methods of analysis (16th ed.). Washington, DC: Association of Official Analytical Chemists.
- Bazaz, M.M., and Keshavanath, P. 1993. Effect of feeding different levels of sardine oil on growth, muscle composition and digestive enzyme activities of mahseer, *Tor khudree*. *Aquaculture*, **115**, 111-119.
- Caballero, M.J., Lopez, G., Socotto, J., Roo, F., J., Izquierd, M.S., and Fernandez, A.J. 1999. Combined effect of lipid level and fish meal quality on liver histology of Gilthead sea braem (*Sparus aurata*). *Aquaculture*, **179**, 277-290.
- Cho, S.H. Lee, S.M., and Lee, J.H., 2005. Effect of dietary protein and lipid levels on growth and body composition of juvenile turbot (*Scophthalmus maximus* L.) reared under optimum salinity and temperature conditions. *Aquaculture Nutrition*, **11**, 235-240.
- Fafioye, O.O., Fagade, S.O., Adebisi, A.A., Jenyo-Oni, and Omoyinmi, G.A.K. 2005. Effects of Dietary Soybeans (*Glycine max* (L.) Merr.) on Growth and Body Composition of African Catfish (*Clarias gariepinus*, Burchell) Fingerlings. *Turkish Journal of Fisheries and Aquatic Science*, **5**, 11-15.
- Fajmonová, E., Zelenka, J., Komprda, T., Kladroba, D., and Šarmanová, I. 2003: Effect of sex, growth intensity and heat treatment on fatty acid composition of common carp (*Cyprinus carpio*) fillets. *Czech Journal of Animal Science*, **48**(2), 85-92.
- Gangadhara, B., Nandeesha, M.C., Varghese, J.J., and Keshavanath, P. 1997. Effect of varying protein and lipid levels on the growth of rohu, *Labeo rohita*. *Asian Fisheries Science*, **10**, 139-147.

- Hossain, M.A., Naher, N., and M. Kamal, M. 1997. Nutrient digestibility coefficients of some plant and animal proteins for rohu (*Labeo rohita*). *Aquaculture*, **151**, 37-45.
- Hu, Y.H., Liu, Y.J., Tian, L.X., Yang, H.J., Liang, G.Y., and Gao, W. 2007. Optimal dietary carbohydrate to lipid ratio for juvenile yellowfin seabream (*Sparus latus*). *Aquaculture Nutrition*, **13**, 291-297.
- Ikeda, A.K., Zuanon, J.A.S., Salaro, A.L., Freitas, M.B.D., Pontes, M.D., Souza, L.S., and Santos, M.V. 2011. Vegetable oil sources in diets for freshwater angelfish (*Pterophyllum scalare*, Cichlidae): growth and thermal tolerance. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, **63**(3), 670-677.
- Ionescu, A., Berza, M., and Banu C. 1992. *Metode și tehnici pentru controlul peștelui și al produselor din pește/Methods and techniques for fish and fishery products control*. „Dunărea de Jos” University of Galati Publishing house Galați.
- Kukačka, V., Chaloupková, L., Fialová, M., Kopp, R., and Mareš, J. 2009. The influence of linseed oil and fish oil supplements to the fatty acid spectrum of common carp (*Cyprinus carpio* L.) muscle. *Acta Universitatis Agriculturae et Silviculturae Mendeleianae Brunensis*, LVII, **5**, 193-192.
- Luo, Z., Liu, Y.J., Mai, K.S., Tian, L.X., Liu, D.H., Tan, X.Y., and Lin, H.Z. 2005. Effect of dietary lipid level on growth performance, feed utilization and body composition of grouper *Epinephelus coioides* juveniles fed isonitrogenous diets in floating netcages. *Aquaculture International*, **13**, 257-269.
- Manjappa, K., Keshavanath, P., and Gangadhara, B. 2002. Growth performance of common carp, *Cyprinus carpio* fed varying lipid levels through low protein diet, with a note on carcass composition and digestive enzyme activity. *Acta Ichthyologica et Piscatoria*, **32**, 146-155.
- Manjappa, K., Keshavanath, P., and Gangadhara, B. 2011. Influence of sardine oil supplemented fish meal free diets on common carp (*Cyprinus carpio* L.) growth, carcass composition and digestive enzyme activity. *Journal of Fisheries and Aquatic Science*, **6**(6), 604-613.
- Martino, R.C., Cyrino, J.E.P., Portz, L., and Trugo, L.C. 2002. Effect of dietary lipid level on nutritional performance of the surubim, *Pseudoplatystoma coruscans*. *Aquaculture*, **209**, 209-218.
- Narejo, N.T., Dars, B.A., and Achakzai, G.D. 2010. Preparation of low-cost fish feed for the culture of *Labeo rohita* (Hamilton) in glass aquaria. *Sindh University Research Journal (Science. Series)*, **42**(2), 7-10.
- Peres, H., and Oliva-Teles, A. 1999. Effect of dietary lipid level on growth performance and feed utilization by European sea bass juveniles *Dicentrarchus labrax*. *Aquaculture*, **179**, 325-334.
- Satpathy, B.B., Mukherjee, D., and Ray, A.K. 2003. Effect of dietary protein and lipid levels on growth, feed conversion on body composition in rohu, *Labeo rohita* (Hamilton), fingerlings. *Aquaculture Nutrition*, **9**, 17-24.
- Steffens, W. 1996. Protein sparing effects and nutritive significance of lipid supplementation in carp diet. *Archives of Animal Nutrition*, **49**, 93-98.
- Steffens, W., and Wirth, M. 2007. Influence of nutrition on the lipid quality of pond fish: common carp (*Cyprinus carpio*) and tench (*Tinca tinca*). *Aquaculture International*, **15**(3-4), 313-319.
- Tuan, L.A., and Williams, K.C. 2007. Optimum dietary protein and lipid specifications for juvenile malabar grouper (*Epinephelus malabaricus*). *Aquaculture*, **267**, 129-138.
- Schulz, C., Huber, M., Ogunji J., and Rennert, B. 2008. Effects of varying dietary protein to lipid ratios on growth performance and body composition of juvenile pike perch (*Sander lucioperca*). *Aquaculture Nutrition*, **14**, 166-173.